

HIZLI ANTİBİYOTİK DUYARLILIK TESTLERİ

Prof. Dr. Gülşen Çetin Hazırolan



İleri Eğitim Programı
Tanısal Yönetişim
5-6.12.2025, Ankara



Antimicrobial Stewardship Core Team

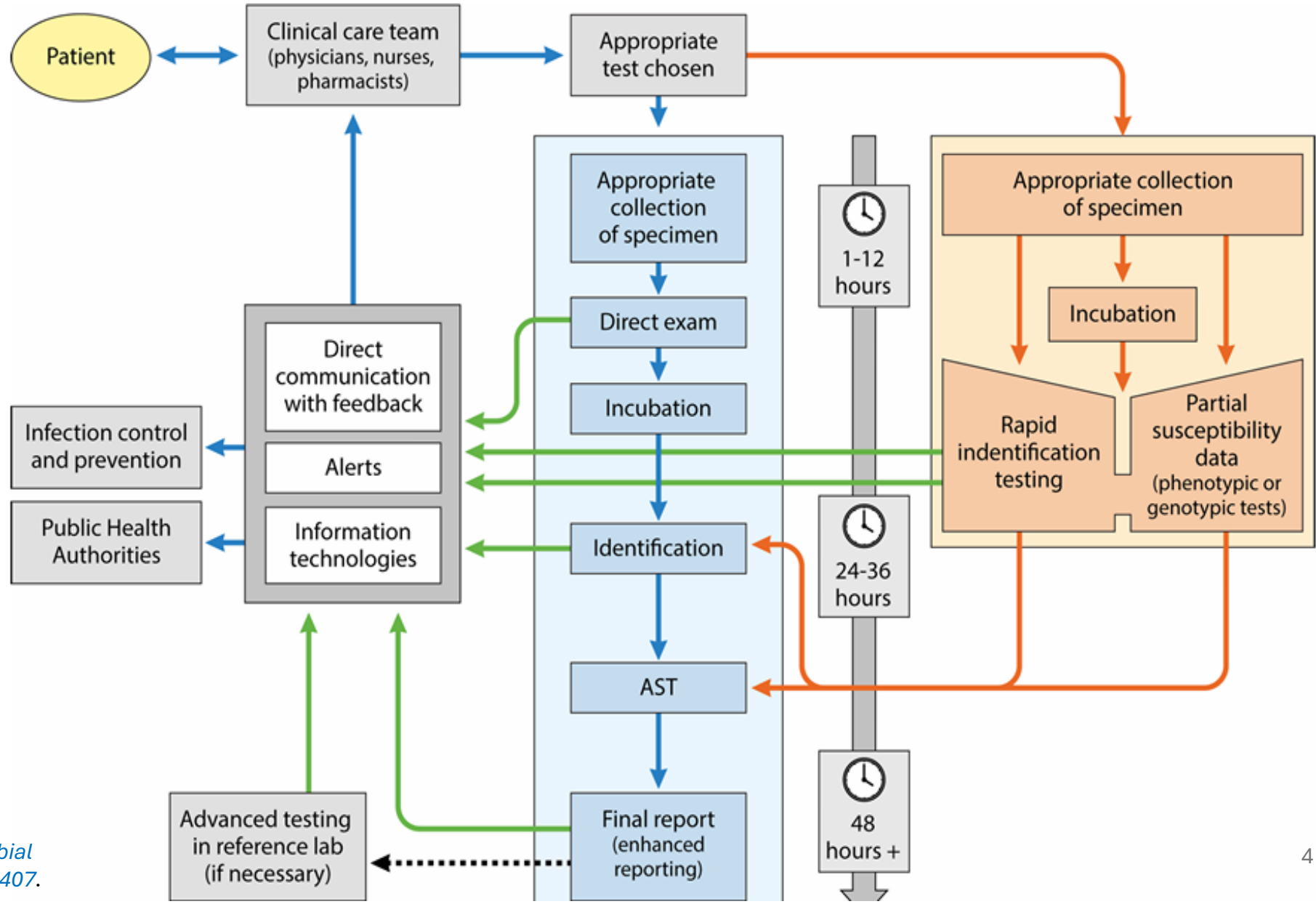




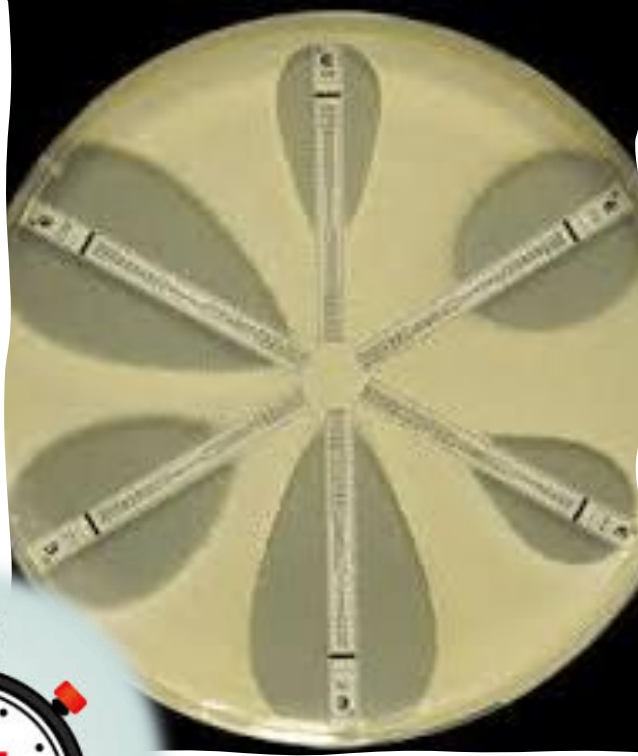
Microbiology lab

- Etkeninin **dođru-hızlı** tanımlanması, antibiyotik duyarlılık test sonuçlarının **standart** şekilde çalışılması, zamanında ve dođru sonucun verilmesi
- Klinisyen etkene özđü tedaviyi seçebilir
- Geniş spektrumlu antibiyotik kullanımının engellenmesinde ve **AMR** mücadelede önemli bir rol oynar

Klinik Mikrobiyoloji Laboratuvarları



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Antibiyotik duyarlılık testleri (ADT)

Kantitatif yöntemler (MİK, mg/L)

- Agar veya broth dilüsyon
- Gradyan şeritleri (Etest, MICE)

Kalitatif yöntemler (S/I/R)

- Disk difüzyon

Otomatize sistemler

- ÇİD-TİD izolatların saptanması
- Doğal Direnç ve Beklenmeyen Fenotip



Rutin ADT metodlarının günler ile ifade edilen süreleri yavaş kalmaktadır...

YENİ VE HIZLI ADT



ADT
sonuç
zamanı



Bildirilen
sonuçların
klinik
değeri



YENİ VE HIZLI ADT

VS

GENOTYPE

PHENOTYPE



GENOTİPİK HIZLI ADT

Önceden tanımlanmış hedefler (direnç mekanizmaları)

Gen VAR- YOK

Duyarlı = tespit edilen direnç mekanizmalarının yokluğu

Asla duyarlılığı garanti edemez, en iyi durumda direnci garanti edebilir

Tespit edilen direnç mekanizmalarının yokluğu, ampirik tedavinin azaltılması için yeterli midir?

GENOTİPİK HIZLI ADT

- Genellikle iyi tanımlanmış direnç mekanizmaları
MRSA/VRE, OXA-48, KPC, NDM, VIM, IMP, mcr
- Özgüllük ve özellikle duyarlılıkları %100 değil
- Çok fazla direnç geni – kompleks direnç mekanizmaları
- Gram (-)'lerde direnç saptayan testler az
- Mutasyonel direnç
- İnd. direnç





Moleküler Testler



Test	MO	Direnç geni
Biofire BCID2 (Biofire, Salt Lake City, UT) 1 s	9 GP -14 GN -7 MAYA	blaIMP, blaKPC,, blaOXA-48-like blaNDM, blaVIM mcr-1 ESBL blaCTX-M mecA/C vanA/B
Verigene BC-GN (Luminex, Austin, TX) 9 Gram-negative bacterial targets	9 GN	blaIMP blaKPC blaOXA-48-like blaNDM blaVIM ESBL blaCTX-M
Verigene BC-GP (Luminex)	13 GP	mecA -vanA/B
ePlex® BCID-GP (GenMark, Carlsbad, CA)	20 GP	mecA -vanA/B
ePlex® BCID-GN (GenMark)	21 GN	blaIMP blaKPC blaOXA-48/OXA-23 blaNDM blaVIM ESBL blaCTX-M
Xpert® MRSA/SA BC (Cepheid, Sunnydale CA)	1 GP	mecA



Acinetobacter calcoaceticus-baumannii complex

Bacteroides fragilis

Enterobacterales

- *Enterobacter cloacae* complex
- *Escherichia coli*
- *Klebsiella aerogenes*
- *Klebsiella oxytoca*
- *Klebsiella pneumoniae* group
- *Proteus* spp.
- *Salmonella* spp.
- *Serratia marcescens*

Haemophilus influenzae

Neisseria meningitidis

Pseudomonas aeruginosa

Stenotrophomonas maltophilia

Candida albicans

Candida auris

Candida glabrata

Candida krusei

Candida parapsilosis

Candida tropicalis

Cryptococcus (C. neoformans/C. gattii)

Enterococcus faecalis

Enterococcus faecium

Listeria monocytogenes

Staphylococcus spp.

- *Staphylococcus aureus*
- *Staphylococcus epidermidis*
- *Staphylococcus lugdunensis*

Streptococcus spp.

- *Streptococcus agalactiae*
- *Streptococcus pneumoniae*

Streptococcus pyogenes

Carbapenemases

IMP

KPC

OXA-48-like

NDM

VIM

Colistin Resistance

mcr-1

ESBL

CTX-M

Methicillin Resistance

mecA/C

mecA/C and MREJ (MRSA)

Vancomycin Resistance

vanA/B



14 G (-)

7 Mantar

9 G (+)

11 Direnç geni



Critically ill;
Drug-resistant
pathogen;
Intensive care units;
Multiplex polymerase
chain reactio



Methods: This retrospective observational study, conducted from July 2021 to August 2023, involved adult ICU patients with positive blood cultures who underwent BCID2 testing. The concordance between BCID2 and conventional culture results was examined, and its impact on antimicrobial stewardship was assessed through a comprehensive retrospective review of patient records by intensivists.

Results: A total of 129 blood specimens from 113 patients were analysed. Among these patients, a high proportion of drug-resistant strains were noted, including carbapenem-resistant *Klebsiella pneumoniae* (CRKP) (57.1%), carbapenem-resistant *Acinetobacter calcoaceticus-baumannii* complex (100%), methicillin-resistant *Staphylococcus aureus* (MRSA) (70%), and vancomycin-resistant *Enterococcus faecium* (VRE) (100%). The time from blood culture collection to obtaining BCID2 results was significantly shorter than conventional culture (46.2 h vs. 86.9 h, $p < 0.001$). BCID2 demonstrated 100% concordance in genotype–phenotype correlation in antimicrobial resistance (AMR) for CRKP, carbapenem-resistant *Escherichia coli*, MRSA, and VRE. A total of 40.5% of patients received inadequate empirical antimicrobial treatment. The antimicrobial regimen was adjusted or confirmed in 55.4% of patients following the BCID2 results.

Conclusions: In the context of a high burden of drug-resistant pathogens, BCID2 demonstrated rapid pathogen and AMR detection, with a noticeable impact on antimicrobial stewardship in BSI in the ICU.

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YBÜ-113 HASTA

%57.1: CRKP

%100: CRABCC

%70 MRSA

%100 VRE

RESEARCH

Clinical impact and cost-consequence analysis of ePlex® blood identification panels for the rapid diagnosis of bloodstream infections: a single-center randomized controlled trial

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M. Maurin^{1,7} · S. David-Tchouda^{9,10,11} · P. Pavese³

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© The Author(s) 2024, corrected publication 2024

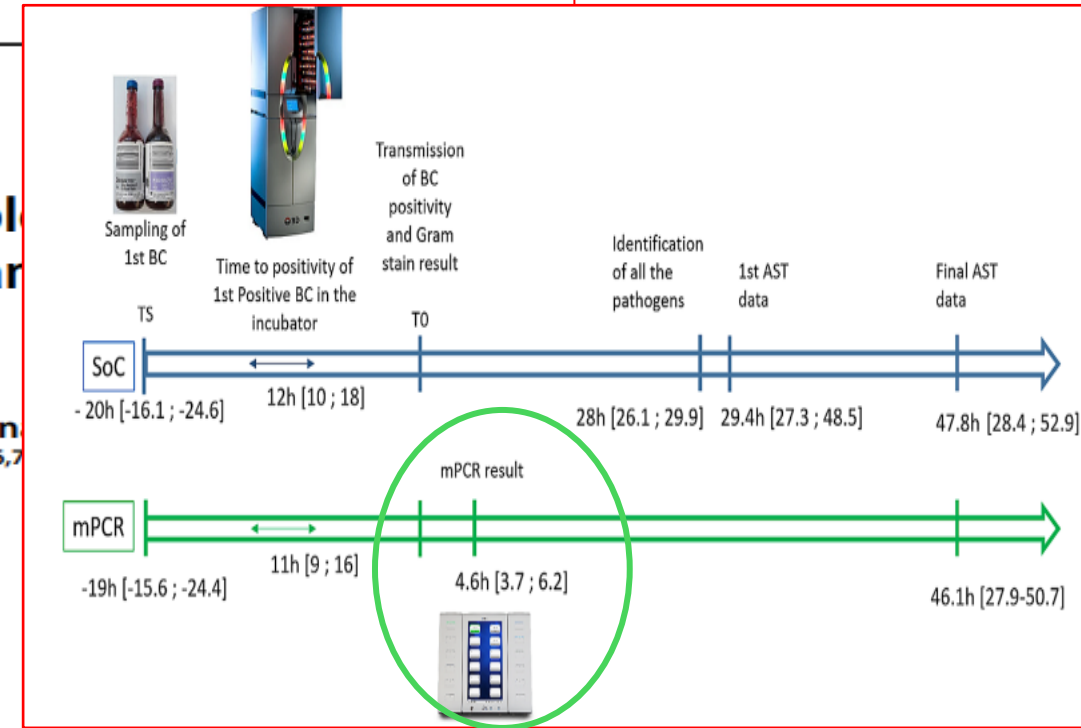
Abstract

Purpose To assess clinical impact and perform cost-consequence analysis of the broadest multiplex PCR panels available for the rapid diagnosis of bloodstream infections (BSI).

Methods Single-center, randomized controlled trial conducted from June 2019 to February 2021 at a French University hospital with an institutional antimicrobial stewardship program. Primary endpoint was the percentage of patients with optimized antimicrobial treatment 12 h after transmission of positivity and Gram stain results from the first positive BC.

Results This percentage was significantly higher in the multiplex PCR (mPCR) group (90/105 = 85.7% %, CI95% [77.5 ; 91.8] vs. 68/107 = 63.6%, CI95% [53.7 ; 72.6]; $p < 10^{-3}$) at interim analysis, resulting in the early termination of the study after the inclusion of 309 patients. For patients not optimized at baseline, the median time to obtain an optimized therapy was much shorter in the mPCR group than in the control group (6.9 h, IQR [2.9; 17.8] vs. 26.4 h, IQR [3.4; 47.5]; $p = 0.001$). Early optimization of antibiotic therapy resulted in a non-statistically significant decrease in mortality from 12.4 to 8.8% ($p = 0.306$), with a trend towards a shorter median length of stay (18 vs. 20 days; $p = 0.064$) and a non-significant reduction in the average cost per patient of €3,065 ($p = 0.15$). mPCR identified all the bacteria present in 88% of the samples.

Conclusion Despite its higher laboratory cost, the use of multiplex PCR for BSI diagnosis leads to early-optimised therapy, seems cost-effective and could reduce mortality and length of stay. Their impact could probably be improved if implemented 24/7.



GENOTİPİK HIZLI ADT

Test maliyeti

X

Antibiyotik kullanım oranı
Tedavinin başlanma süresi
Tedavi süresi
Yatış süresi
Mortaliteye

Yeni Fenotipik ADT

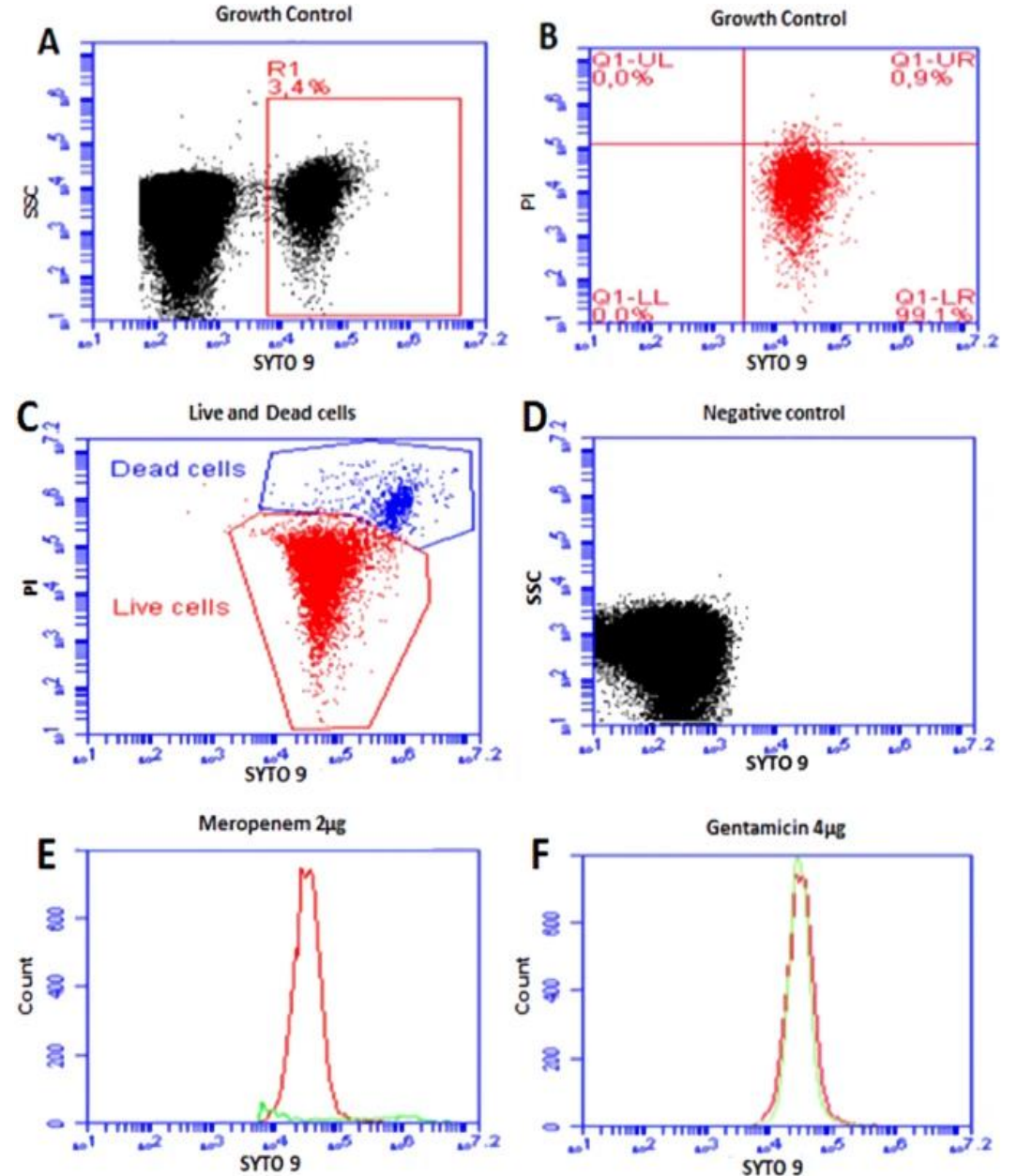
- Kısa süreli antibiyotik maruziyeti
- Antibakteriyel etkilerin yüksek duyarlılıkta tespiti

Popülasyon düzeyinde

- ✓ Flow sitometrisi – hücre sayısı
- ✓ Raman spektroskopisi
- ✓ Mikrok calorimetri
- ✓ Ortalama mo kütlesi

Tek hücre düzeyinde

- ✓ Flow sitometrisi – hücre morfolojisi/floresan
- ✓ Zaman aralıklı mikroskopi – morfoloji/biyokütle



KAN KÜLTÜRLERİNDEN DOĞRUDAN HADT



EN ZOR

EN KOLAY

EN ÇOK İHTİYAÇ
DUYULAN

KAN
ALIMI

KK
POZİTİF

BAKTERİ
ID

ADT



PCR/HİBRİDİZASYON

KAN KÜLTÜRLERİNDEN DOĞRUDAN HADT



TEST	ADT TEKNOLOJİ	TTR	ONAY
PhenoTest BC (Accelerate Diagnostics)	Karanlık alan mikroskopisi ile zaman aralıkları görüntüleme (Bakterideki morfolojik ve kinetik değişiklikleri analiz etme)	7 s	US FDA CE-IVD
dRAST (QuantaMatrix)	Bakterileri hücrelerini plastik mikroçiplerde zaman aralıklı görüntüleme	6s	CE-IVD
Alfred (AliFAX)	Sıvı besiyerinde bakteri üremesini ışık saçma ile tespiti	3-5 s	CE-IVD
Reveal AST (Specific Diagnostics)	Mikroorganizma büyümesi sırasında yayılan uçucu organik bileşikler için sensör dizisi	4-5 s	CE-IVD
ASTar (Q-linea)	Sıvı besiyerinde bakteri üremesinin zaman aralıklı görüntülenmesi	3-6 s	CE-IVD
Fastinov	Akım sitometri :Ab altında oluşan hücre hasarını flörosans boyala ile tespiti	80 dk	CE-IVD

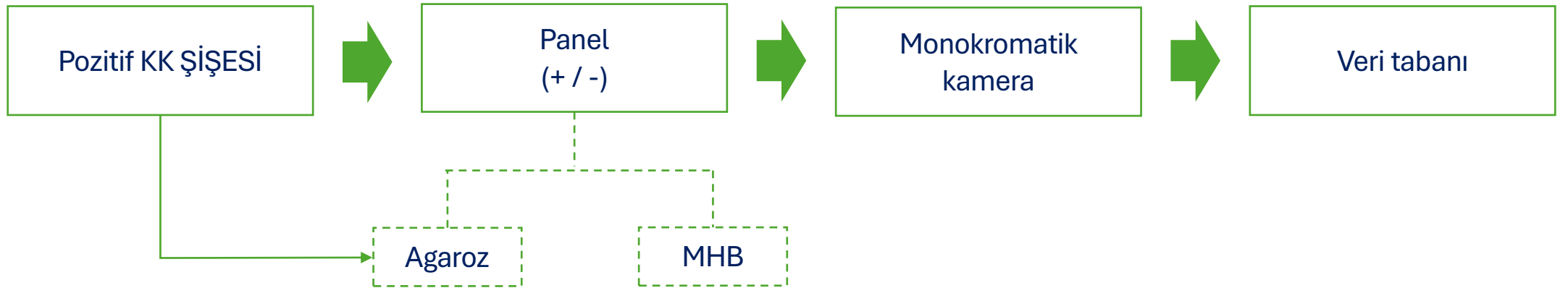
dRAST
Mikrobiyal İyileştirme ve Kontrol Sistemleri

dRAST™

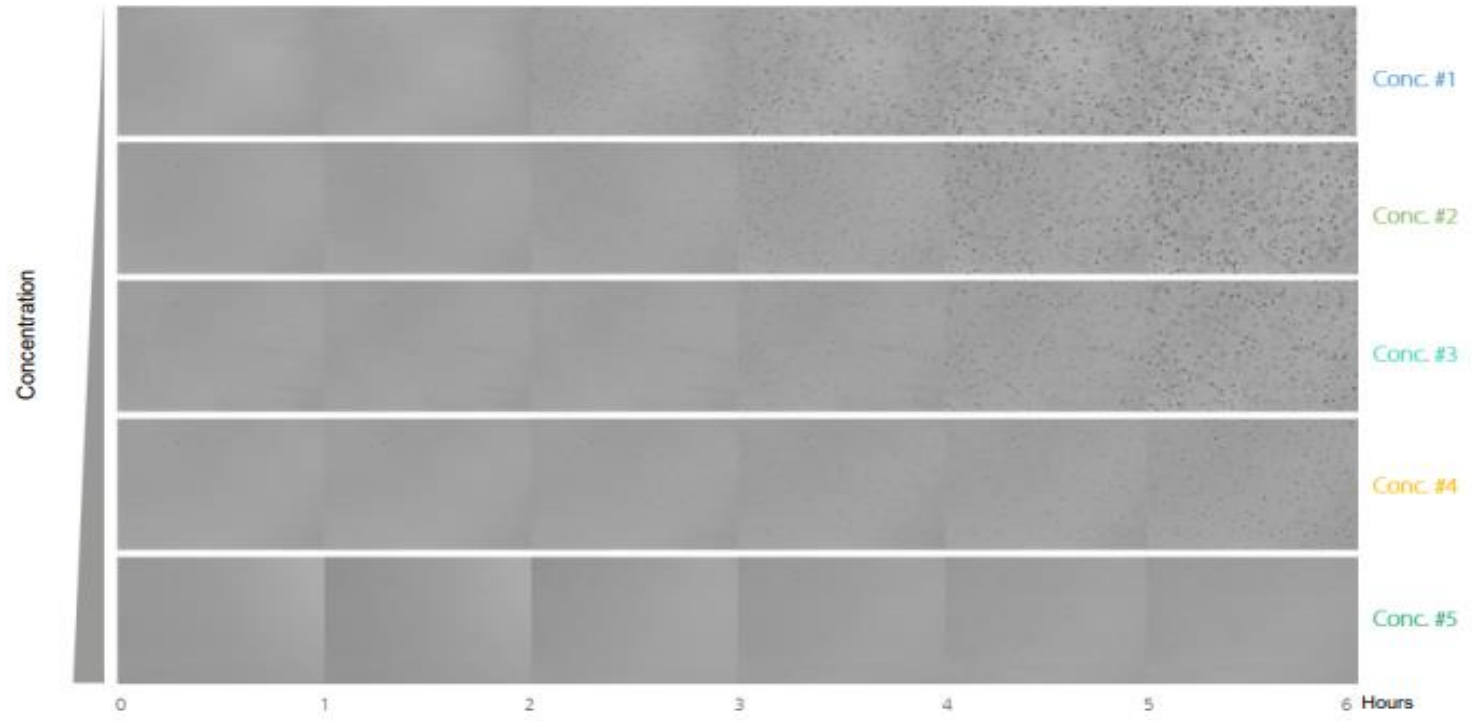
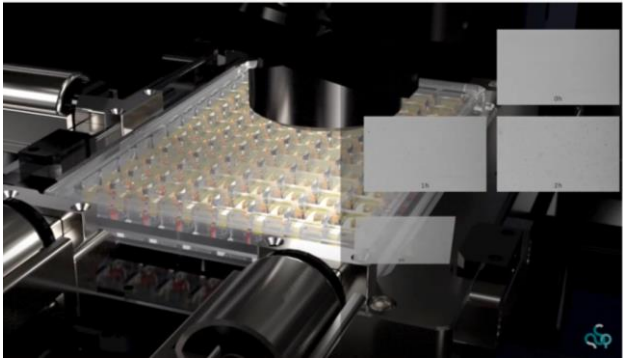
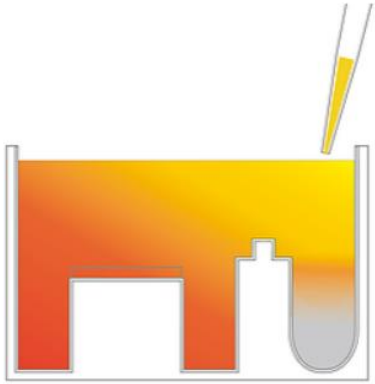
Farklı
konsantrasyonlarda
AB içeren 96
kuyucuklu plak

Yüksek
çözünürlüklü
monokromatik
kamera ile saat
başı çekim (4-6)

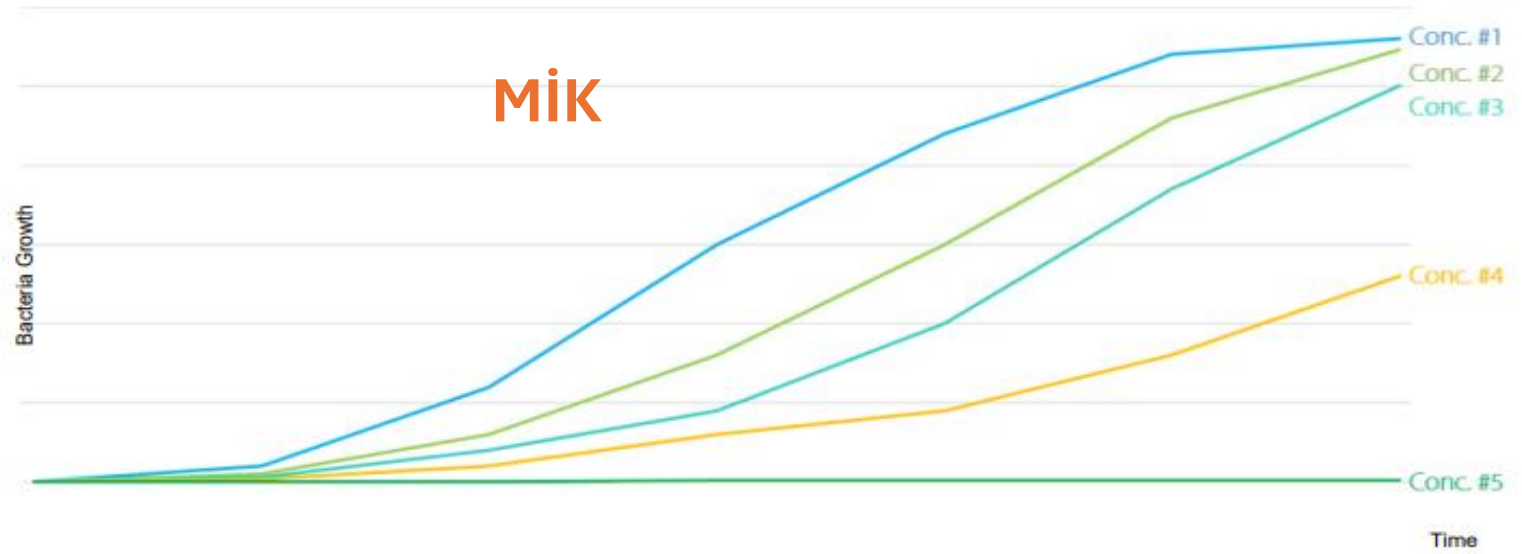
Veri tabanındaki
algoritmalar MİK belirler-
EUCAST kriterlerini
uygulayarak sonuç verir

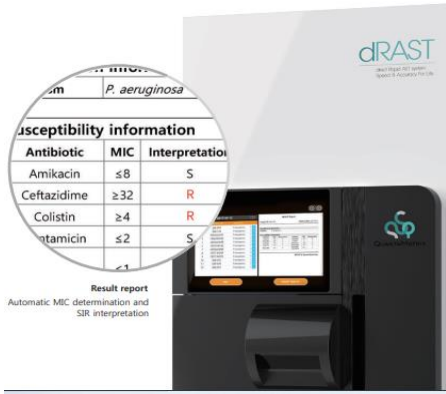


dRAST™



MiK





dRAST™



Performance of dRAST™ on prospective clinical blood culture samples in a simulated clinical setting and on multidrug-resistant bacteria

Alicia Y. W. Wonga , Alexander T. A. Johnssonb , Volkan Özenci

KU: %95.6

MDR-KU: %87.2

Hızlı antibiyotik duyarlılık testi dRAST™ performansının sınırda direnç gösteren mikroorganizmalarda değerlendirilmesi

Gülşen Hazırolan, Onur Karatuna, Volkan Özenci

KU: %95.3

VRE-MRSA-KU: %100

Pa-KU: % 81.3

Direct rapid antibiotic susceptibility test (dRAST) for blood culture and its potential usefulness in clinical practice

Jeong-Han Kim,¹† Taek Soo Kim,²† Sang Hoon Song,² Jungil Choi,³ Sangkwon Han,³ Dong Young Kim,³ Sunghoon Kwon,³ Eunyong Lee,¹ Kyoung-Ho Song,¹ Pyeong Gyun Choe,¹ Ji Hwan Bang,¹ Eu Suk Kim,¹ Sang Won Park,¹ Hong Bin Kim,¹ Nam Joong Kim,¹ Wan Beom Park^{1,*} and Myoung-don Oh¹

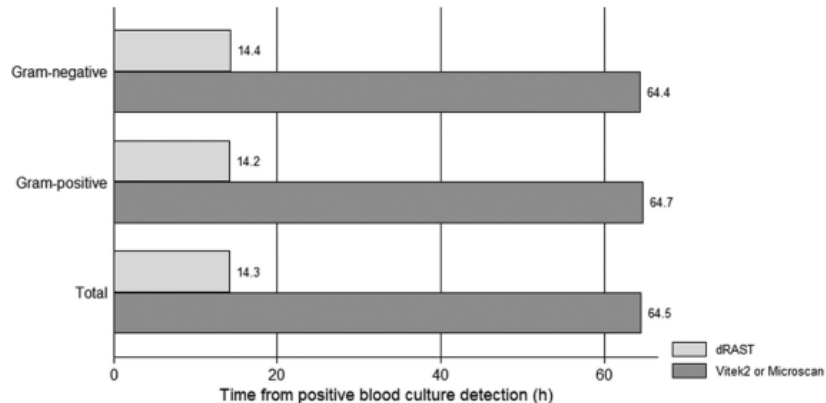
Abstract

Purpose. The direct rapid antibiotic susceptibility test (dRAST), based on analysing changes in bacterial micro-colonies under antibiotic conditions, detects antibiotic resistance within 6 h of direct smear examination results. This study aimed to assess the accuracy of dRAST and evaluate its potential usefulness for improving selection of appropriate antibiotic in real clinical practice settings.

Methodology. We evaluated the accuracy of dRAST by comparing the antibiotic treatments that should have been administered based on dRAST results and the broth microdilution (BMD) test and its potential usefulness via simulation.

Result. For 49/52 (94.2%) patients with Gram-positive bacteraemia and 66/67 (98.5%) patients with Gram-negative bacteraemia, antibiotics indicated by dRAST results were the same as those indicated by the BMD test. Among 34 patients with ineffective and suboptimal treatment, 19 (55.9%) of patients could have received optimal treatment 1 to 2 days earlier with dRAST results. Among 33 patients given unnecessary broad-spectrum antibiotics, 1 to 2 days earlier de-escalation could have been possible for 27 (81.8%) patients based on dRAST results.

Conclusion. The introduction of dRAST could increase the use of optimal antibiotics and reduce unnecessary broad-spectrum antibiotic use in the early period of bacteraemia.

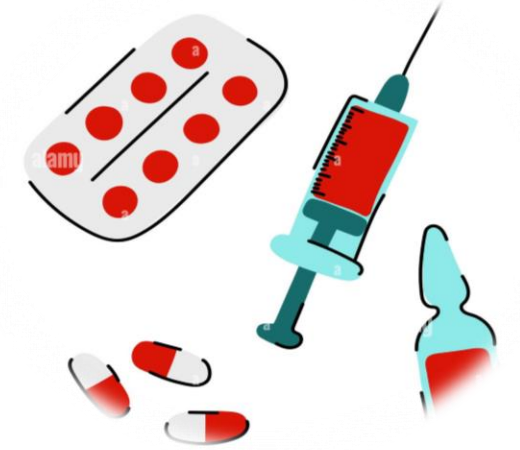


14.3±4.2 - 64.5 ±22.3 s

BMD

%94.2 GP bakteriyemi

%98.5 GN bakteriyemi



Geniş spekt. tdv. % 81.8, DE-ESKELASYON: 1-2 gün

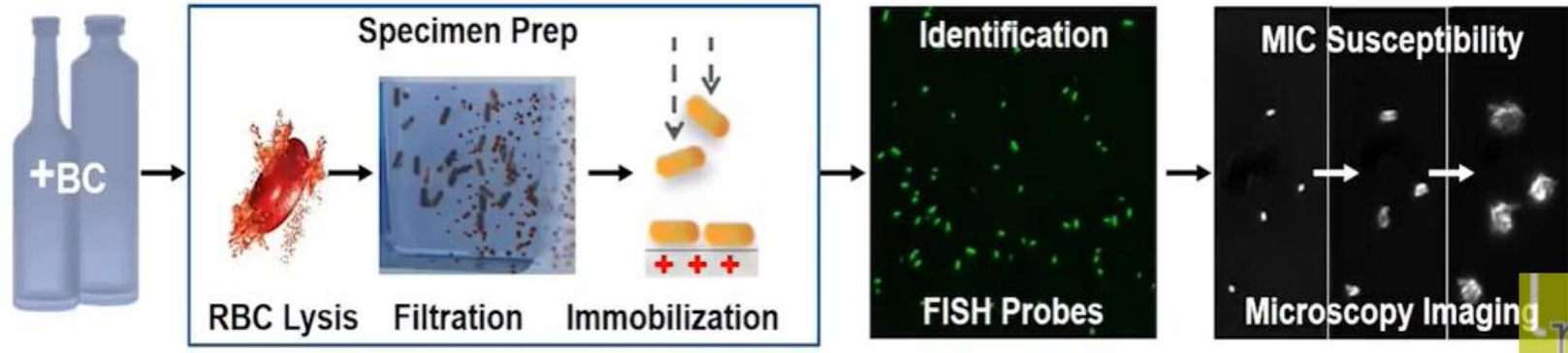
Uygun olmayan tedavi: %55- OPTİMAL TDV: 1-2 gün

Accelerate Pheno System

FISH prob ID (17 mo) -Zaman aralıklı görüntüleme ADT

FDA onaylı

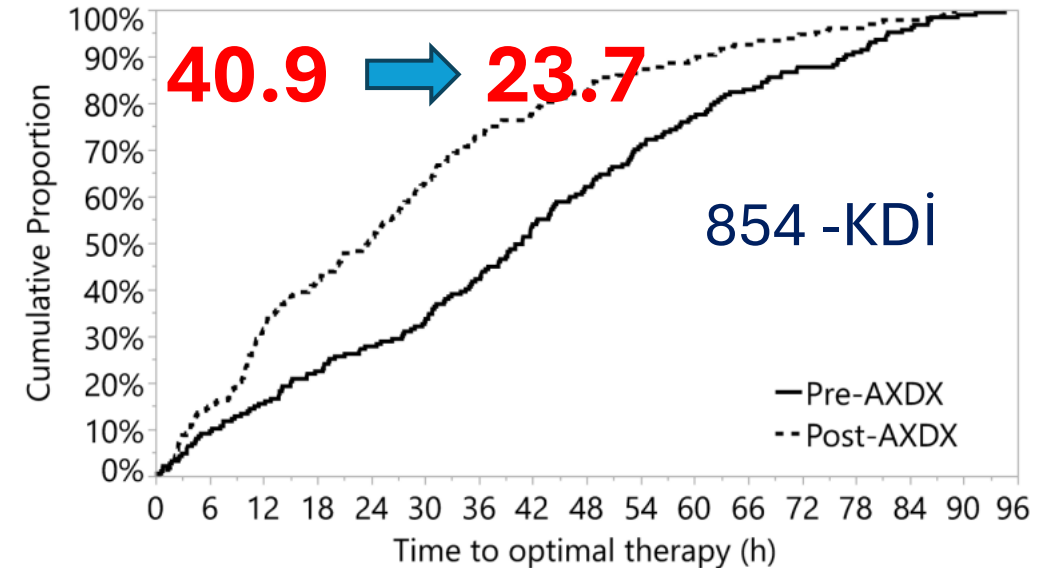
ID: 1.5 s
ADT: <7 s



17 AB ve fenotipik
MRSA & MLSb

Table 4. Antimicrobial Modifications and Clinical Outcomes

Endpoint	All ^a			Gram-Negative ^b		
	Pre-AXDX	Post-AXDX	PValue	Pre-AXDX	Post-AXDX	PValue
Antimicrobial modification^c						
Time to first antimicrobial modification ^d	24.2 (7.3–46.2)	13.9 (5.0–31.1)	<.0001	22.8 (7.0–45.3)	13.6 (5.8–30.9)	.01
Time to first gram-positive antimicrobial modification ^e	30.1 (11.2–52.8)	18.3 (6.7–41.8)	.0013	28.1 (10.5–51.7)	18.6 (9.4–42.1)	.11
Time to first gram-negative antimicrobial modification ^f	34.6 (9.2–53.4)	18.6 (8.2–36.8)	<.0001	30.2 (7.6–52.8)	16.7 (8.6–35.2)	.003
Time to first antimicrobial escalation ^g	9.5 (3.4–28.9)	9.0 (3.7–18.4)	.22	9.5 (3.7–31.6)	9.6 (3.9–18.4)	.44
Time to first antimicrobial deescalation ^h	36.0 (17.1–54.5)	27.2 (13.5–43.6)	.0004	34.5 (16.6–52.8)	25.4 (12.0–42.5)	.003
Time to effective therapy ⁱ	13.3 (3.1–35.9)	6.7 (3.1–16.2)	.02	13.7 (3.3–38.1)	10.0 (3.6–18.6)	.10
Clinical outcome						
30-day mortality	38 (8.7)	25 (6.0)	.12	25 (8.3)	19 (6.7)	.47
Post-blood culture length of stay, median (interquartile range), days	7.0 (4.0–12.4)	6.5 (3.7–12.0)	.43	6.4 (3.7–11.7)	5.4 (3.4–9.7)	.03
Acute kidney injury (aged ≥18 years)	92 (23.2)	78 (21.1)	.49	64 (22.7)	57 (21.6)	.76
14-day renal replacement therapy	15 (3.5)	9 (2.2)	.25	10 (3.3)	5 (1.8)	.24
30-day <i>Clostridioides difficile</i> infection (day 3–30)	3 (0.7)	4 (1.0)	.67	0	1 (0.4)	.48
Acquisition of new multidrug-resistant organisms within 30 days	22 (5.1)	15 (3.6)	.29	17 (5.7)	9 (3.2)	.15
Readmission within 30 days	76 (19.4)	91 (23.8)	.14	52 (18.6)	51 (19.4)	.82
Readmission within 30 days from bacteremia	15 (3.8)	16 (4.2)	.68	7 (2.5)	11 (4.2)	.54



Bhalodi AA et al., Clin Infect Dis. 2021 Oct 27:ciab921.

**KAN
KÜLTÜRLERİNDEN
DOĞRUDAN ADT**

MALDI TOF MS



HIZLI ADT - MALDI-TOF MS

Karakteristik bir “direnç tepe paterni” analizi

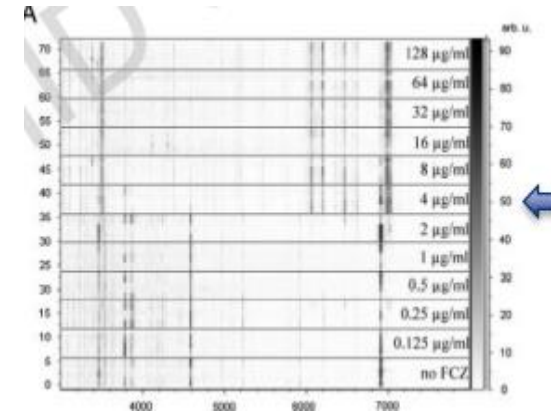
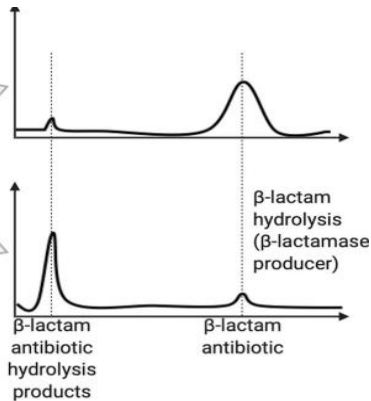
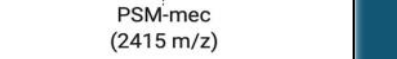
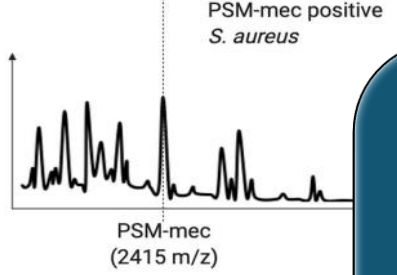
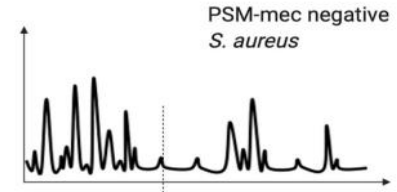
Yeni sentezlenmiş bakteri proteinlerine dâhil edilen kararlı (radyoaktif olmayan) izotop etiketli amino asitlerin saptanması

DİRENÇ TESPİTİ

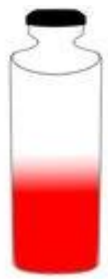
Bakterinin β -laktam antibiyotiği hidroliz analizi

Ab varlığında veya yokluğunda bakteri üremesinin analizi

- β -laktamaz aktivitesini tespiti
- Porin tespiti
- VRE
- MRSA-MSSA
- *B. fragilis* KARBA R



HIZLI ADT - MALDI-TOF MS



Filtrasyon/dilüsyon



Dilüsyon



Lizis/santrifügasyon



Ayırıcı santrifügasyon

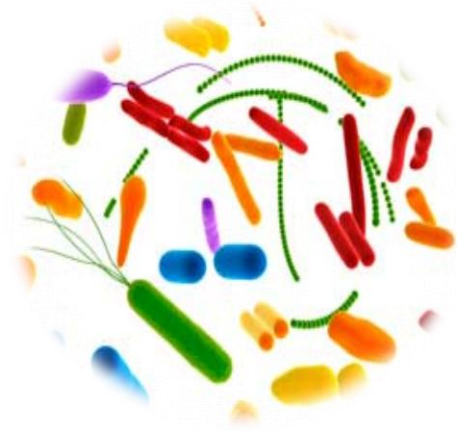
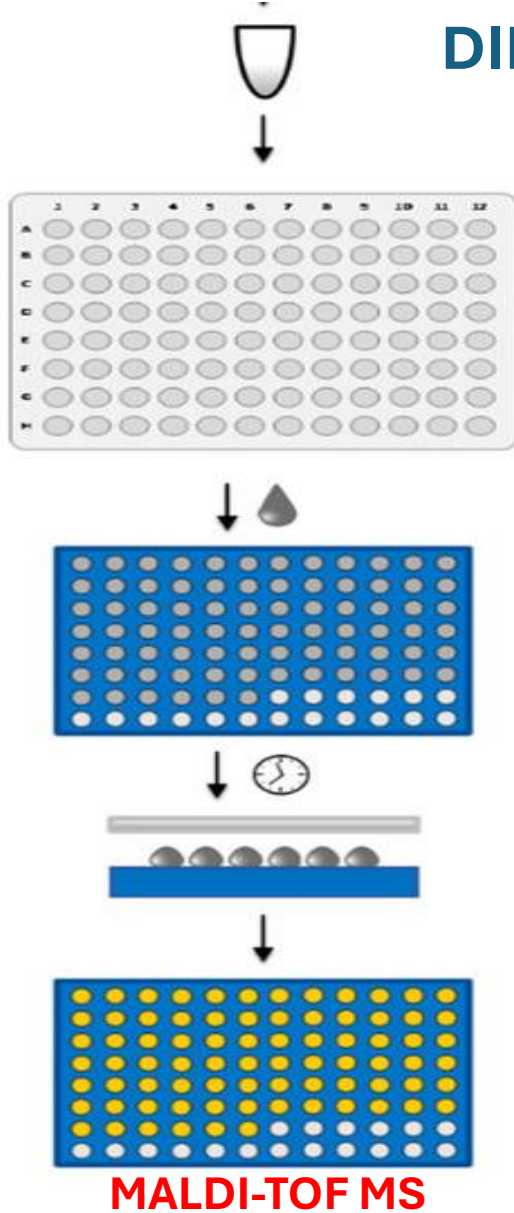


TABLE 2 Performance of MALDI-TOF MS DOT-MGA for direct detection of carbapenem nonsusceptibility in *Enterobacterales* from positive blood cultures using evaluation with MALDI Biotyper scores^a

Processing method	Dilution	Values for 3-h Incubation (%)			Values for 4-h Incubation (%) ^b		
		Validity ^c	Sensitivity ^d	Specificity ^d	Validity	Sensitivity	Specificity
Filtration/dilution	1:100	96.3	100	42.9	100	92.3	14.3
	1:1,000	96.3	91.7	71.4	100	84.6	64.3
	1:10,000	85.2	80.0	100	88.9	81.8	100
Dilution	1:100	100	92.3	71.4	96.3	100	46.2
	1:1,000	96.3	83.3	85.7	100	84.6	92.9
	1:10,000	92.6	63.6	100	92.6	90.9	100
Lysis/centrifugation		92.6	90.9	100	96.3	91.7	100
Differential centrifugation		92.6	90.9	92.9	96.3	83.3	100

DIRECT-ON-TARGET MICRODROPLET GROWTH ASSAY

DOT-MGA



MALDI-TOF MS

Bakteri Ab varlığı ve yokluğunda (ÜK) ink.

Farklı Ab kont-seri dilüsyonlar- Hedef MİK

Mikrodamlacıklar direkt plağa sürülür

4 ml su içeren plastik kutularda ink.

Absorban ile broth bakteri-ab karışımından uzaklaştırılır.

MALDI TOF MS: Üreme var/yok

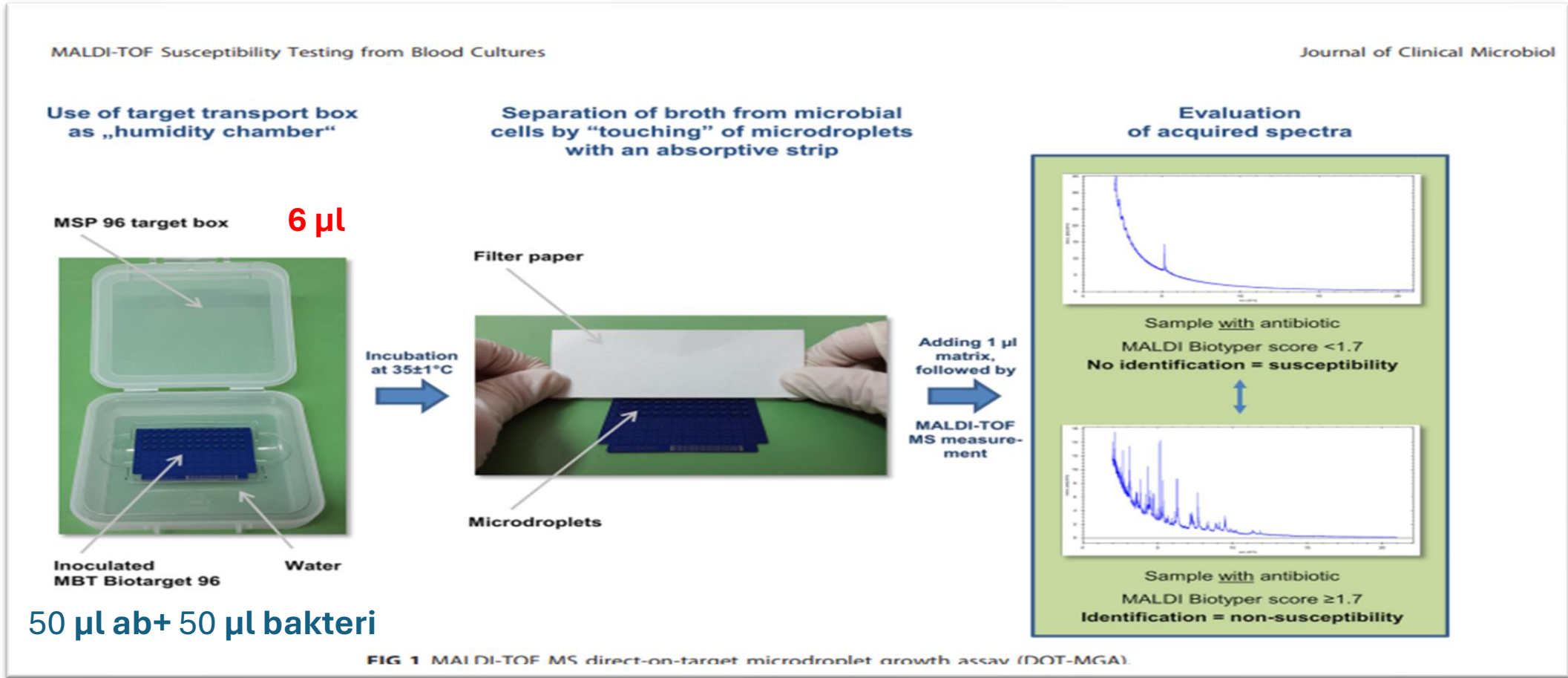
Plaktaki her kuyucukta AB kons: MİK



Rapid Direct Susceptibility Testing from Positive Blood Cultures by the Matrix-Assisted Laser Desorption Ionization–Time of Flight Mass Spectrometry-Based Direct-on-Target Microdroplet Growth Assay

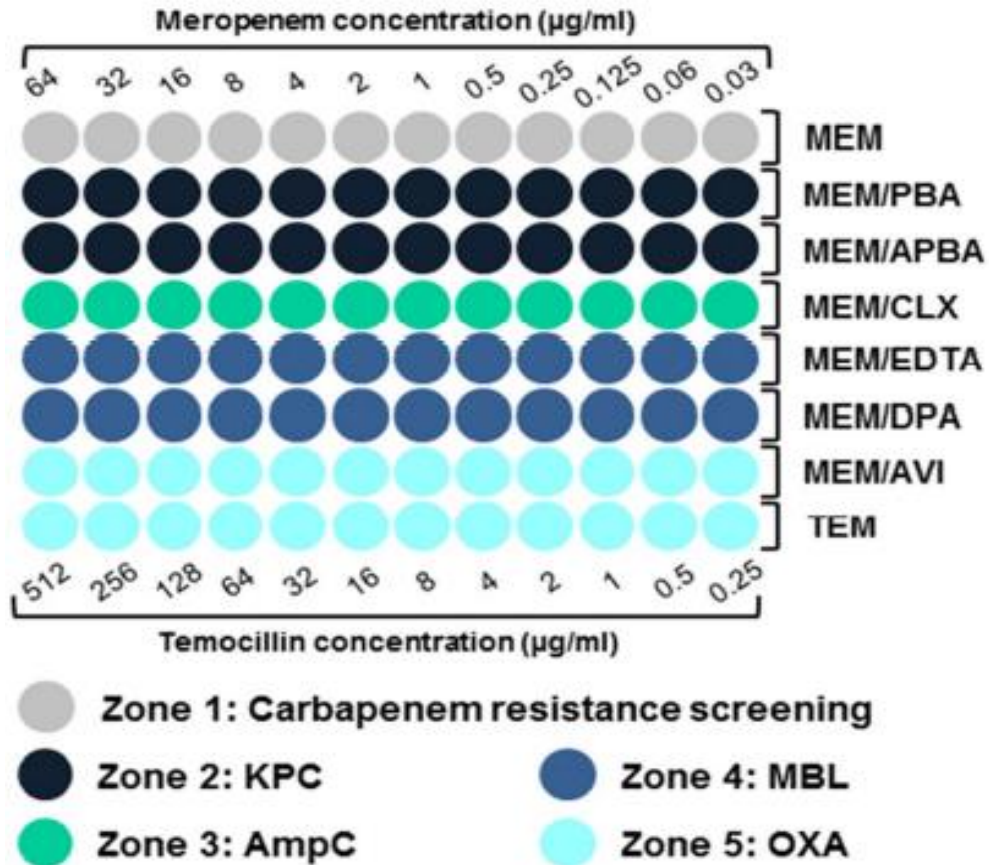
Evgeny A. Idelevich,^a Luise M. Storck,^a Katrin Sparbier,^b Oliver Drews,^b Markus Kostrzewa,^b Karsten Becker^a

Skor ≤ 1.7 NS
Skor ≥ 1.7 S



3 -4 s at 35 \pm 1 $^{\circ}$ C.

MALDI-TOF MS & DOT-MGA

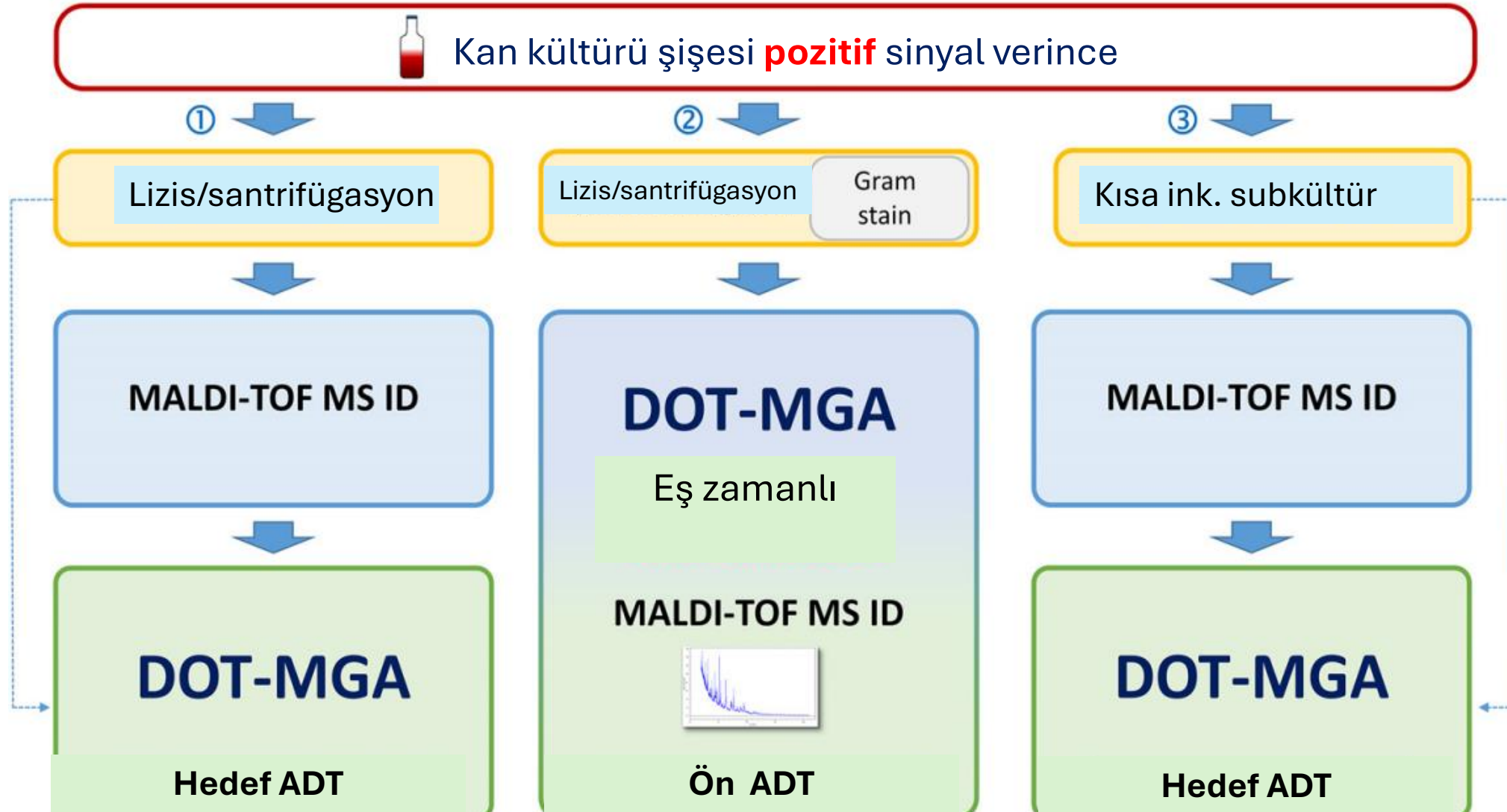


MİK TESPİTİ

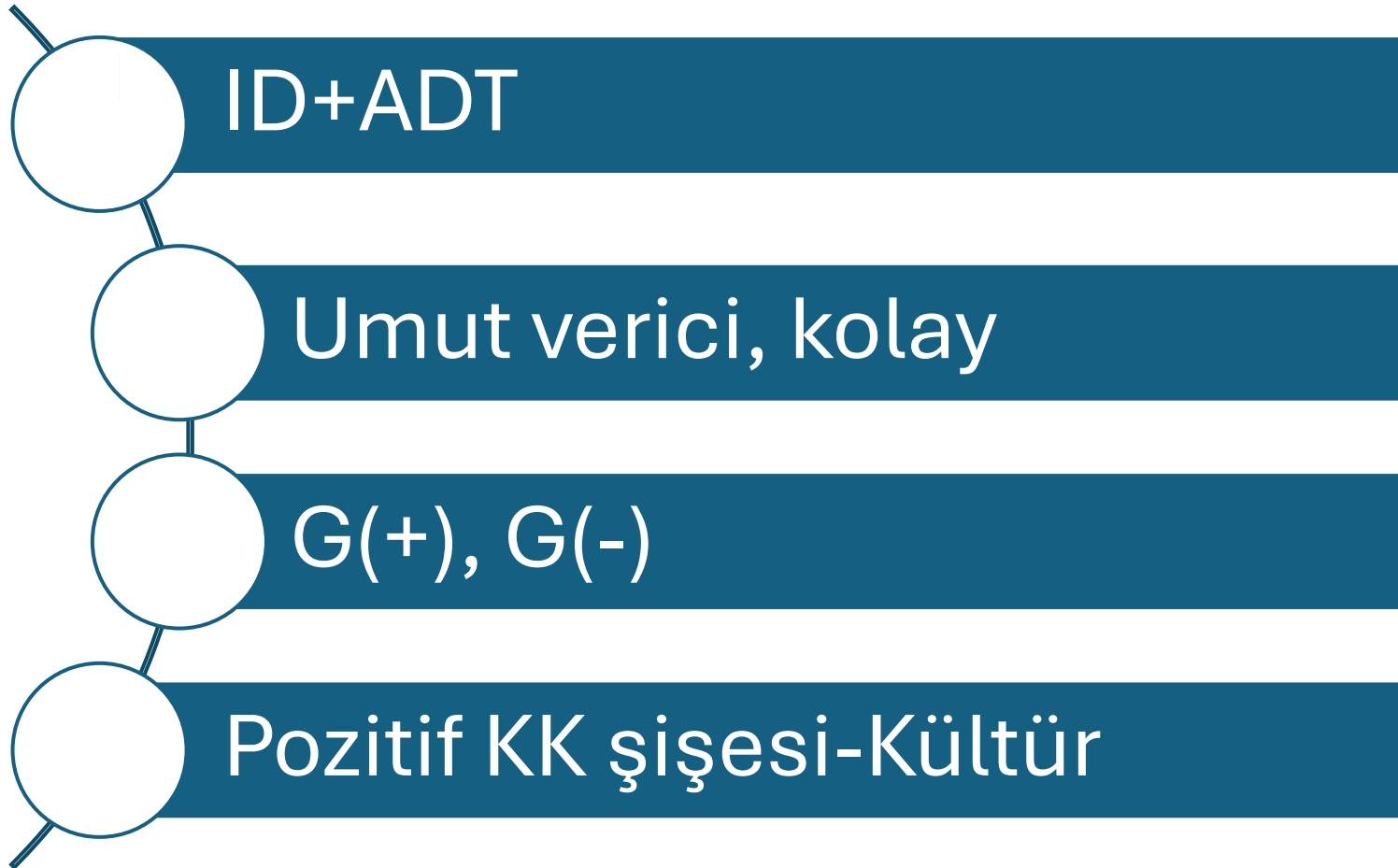
ADT sonucu : 4- 5 s (pozitif KK den)

5×10^5 CFU/ml: 3 µl
3 µl AB

MALDI TOF MS HIZLI TANIMLAMA ve ADT



MALDI-TOF MS & DOT-MGA





Ülkemiz-HADT.....

Methodology - EUCAST rapid antimicrobial susceptibility testing (RAST)
directly from positive blood culture bottles.

Version 1.1

May 2019

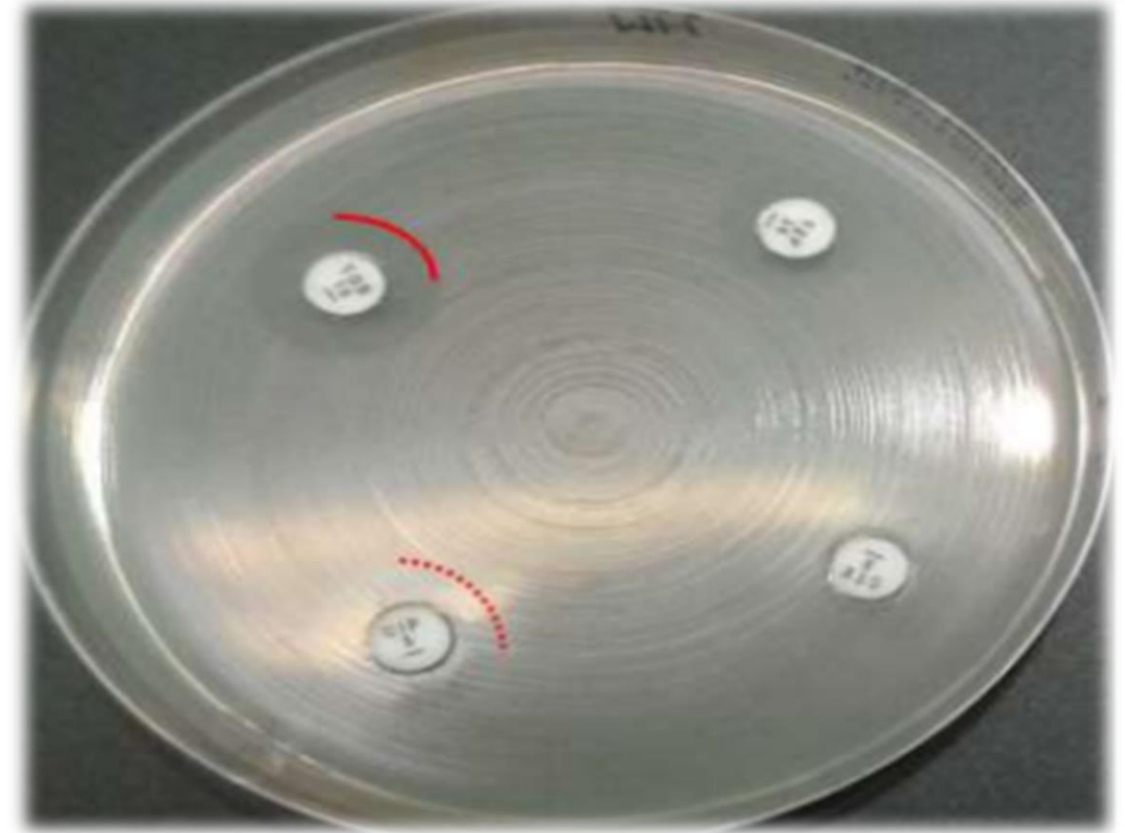


STANDART METODOLOJİ, MATERYAL VE EKİPMAN KULLANARAK....

- ✓ Tüm lab. uygulanabilir...
- ✓ En önemli bakteriyel etkenler...
- ✓ Sepsis tdv. kullanılan ab...
- ✓ Basit işlemler...
- ✓ Kalibre direnç sınır değerleri...
- ✓ Web sitesinde kullanıma açık...

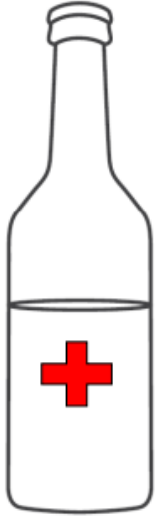
Kan Kültürü Şişelerinden Doğrudan Hızlı Antibiyotik Duyarlılık Testi (HADT)

- Her MH/MH-F agar plağı için 100-150 µl seyreltilmemiş kan kültürü besiyeri, pozitif kan kültürü şişesinden doğrudan alınır
- 4, 6, 8, 16-20 saat
- *E. coli*
- *K. pneumoniae*
- *Salmonella enterica*
- PA, ABCC
- SA, *S. pneumoniae*
- *E. faecium/faecalis*



4 saatlik inkübasyon sonunda *E. coli*.

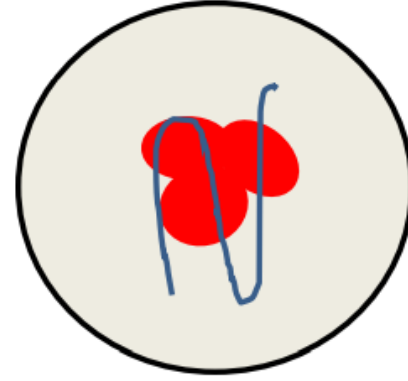
İNOKÜLASYON



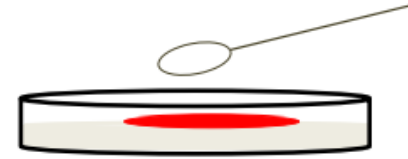
Gram/ID 3 s



125±25 µL



Dağıt

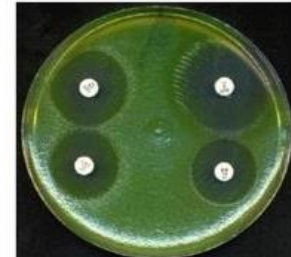
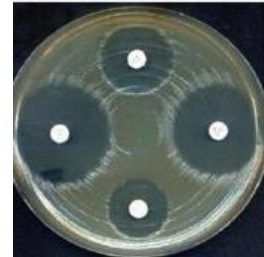


Yay

18 s





Maksimum 4-6 disk



OKUNABİLİRLİK...



	4 s (%)	6 s (%)	8 s (%)	16-20 s (%)
<i>Escherichia coli</i>	90	99	99	100
<i>Klebsiella pneumoniae</i>	96	98	98	100
<i>Pseudomonas aeruginosa</i>	 -	88	97	100
<i>Acinetobacter baumannii</i>	99	100	100	100
<i>Staphylococcus aureus</i>	 55	91	95	100
<i>Enterococcus faecalis</i>	93	99	100	100
<i>Enterococcus faecium</i>	44	93	99	100
<i>Streptococcus pneumoniae</i>	68	83	95	100

Klebsiella pneumoniae

Zone diameter breakpoints for RAST directly from blood culture bottles

EUCAST RAST Breakpoint Tables v. 8.1, valid from 2025-07-14

EUCAST rapid disk diffusion method directly from positive blood culture bottles

Medium: Mueller-Hinton (MH) agar

Inoculum: 125±25 µL directly from a positive blood culture bottle

Incubation: Air, 35±1°C

Incubation time: 4, 6, 8 and 16-20 hours

General reading instructions: Inhibition zones should only be read when the growth is confluent and zone edges are clearly visible.

Reading 4, 6 and 8 hours: Remove the lid and read zone diameters from the front against a dark background illuminated with reflected light.

Reading 16-20 hours: Read zone diameters from the back of the plate against a dark background illuminated with reflected light.

[QC for implementation of RAST](#)

Breakpoints are valid for *K. pneumoniae*, *K. variicola* and *K. quasipneumoniae*.

Antimicrobial agent	Disk content (µg)	4 hours			6 hours			8 hours			16-20 hours		
		S ≥	ATU	R <	S ≥	ATU	R <	S ≥	ATU	R <	S ≥	ATU	R <
Amoxicillin-clavulanic acid	20-10	15	13-14	13	16	14-15	14	16	14-15	14	18	16-17	16
Piperacillin-tazobactam	30-6	15	13-14	13	16	14-15	14	16	14-15	14	17	15-16	15
Temocillin	30	50	14	14	50	15	15	50	16	16	50	16	16
Cefotaxime ¹	5	15	12-14	12	18	15-17	15	18	15-17	15	16	14-15	14
Ceftazidime ¹	10	15	13-14	13	16	14-15	14	16	14-15	14	18	15-17	15
Ceftazidime-avibactam	10-4	12	10-11	10	13	11-12	11	13	11-12	11	14	12-13	12
Ceftolozane-tazobactam	30-10	16	14-15	14	16	14-15	14	17	15-16	15	20	17-19	17
Imipenem ²	10	16	14-15	14	17	15-16	15	17	15-16	15	15	12-14	12
Imipenem-relebactam	10-25	15	13-14	13	15	14	14	15	14	14	17	15-16	15
Meropenem ²	10	15	13-14	13	17	15-16	15	17	15-16	15	15	13-14	13
Meropenem-vaborbactam	20-10	16	14-15	14	17	16	16	17	16	16	15	13-14	13
Ciprofloxacin	5	17	15-16	15	18	16-17	16	18	16-17	16	19	17-18	17
Levofloxacin	5	17	14-16	14	18	15-17	15	18	15-17	15	18	15-17	15
Amikacin ³	30	(15)	(13-14)	(13)	(14)	(12-13)	(12)	(14)	(12-13)	(12)	(15)	(13-14)	(13)
Gentamicin ³	10	(14)	(12-13)	(12)	(14)	(12-13)	(12)	(13)	(11-12)	(11)	(14)	(13)	(13)
Tobramycin ³	10	(14)	(12-13)	(12)	(13)	(11-12)	(11)	(13)	(11-12)	(11)	(14)	(13)	(13)
Trimethoprim-sulfamethoxazole	1.25-23.75	11	9-10	9	11	9-10	9	11	9-10	9	10	8-9	8

AB seçimi: Gram negatif

MİKROORGANİZMA

ANTİBİYOTİK

Escherichia coli
Klebsiella pneumoniae

Piperacillin-tazobactam, Ceftazidime, **Cefotaxime**, Ceftazidime-avibactam, Ceftolozane-tazobactam, Imipenem, **Meropenem**, **Ciprofloxacin**, Levofloxacin, Amikacin, Gentamicin, Tobramycin, Trimethoprim-sulfamethoxazole
Ampicillin • Amoxicillin-clavulanic acid • Temocillin • Imipenem-relebactam • Meropenem-vaborbactam

Pseudomonas aeruginosa

Piperacillin-tazobactam, Cefepime, Ceftazidime, Ceftazidime-avibactam, Ceftolozane-tazobactam, Imipenem, Meropenem, Ciprofloxacin, Levofloxacin, Amikacin, Tobramycin
Imipenem-relebactam • Meropenem-vaborbactam

Acinetobacter baumannii

Imipenem, Meropenem, Ciprofloxacin, Levofloxacin, Amikacin, Gentamicin, Tobramycin, Trimethoprim-sulfamethoxazole



AB seçimi: Gram pozitif

MİKROORGANİZMA

ANTİBİYOTİK

Staphylococcus aureus

Cefoxitin (TARAMA), Norfloxacin (TARAMA), Amikacin, Gentamicin, Tobramycin, Clindamycin

Enterococcus faecalis
Enterococcus faecium

Ampicillin, Imipenem, Vancomycin, Linezolid, Gentamicin (YÜKSEK DÜZEY AG DİRENCİ)

Streptococcus pneumoniae

Oxacillin (TARAMA), Norfloxacin (TARAMA), Erythromycin, Clindamycin, Trimethoprim-sulfamethoxazole

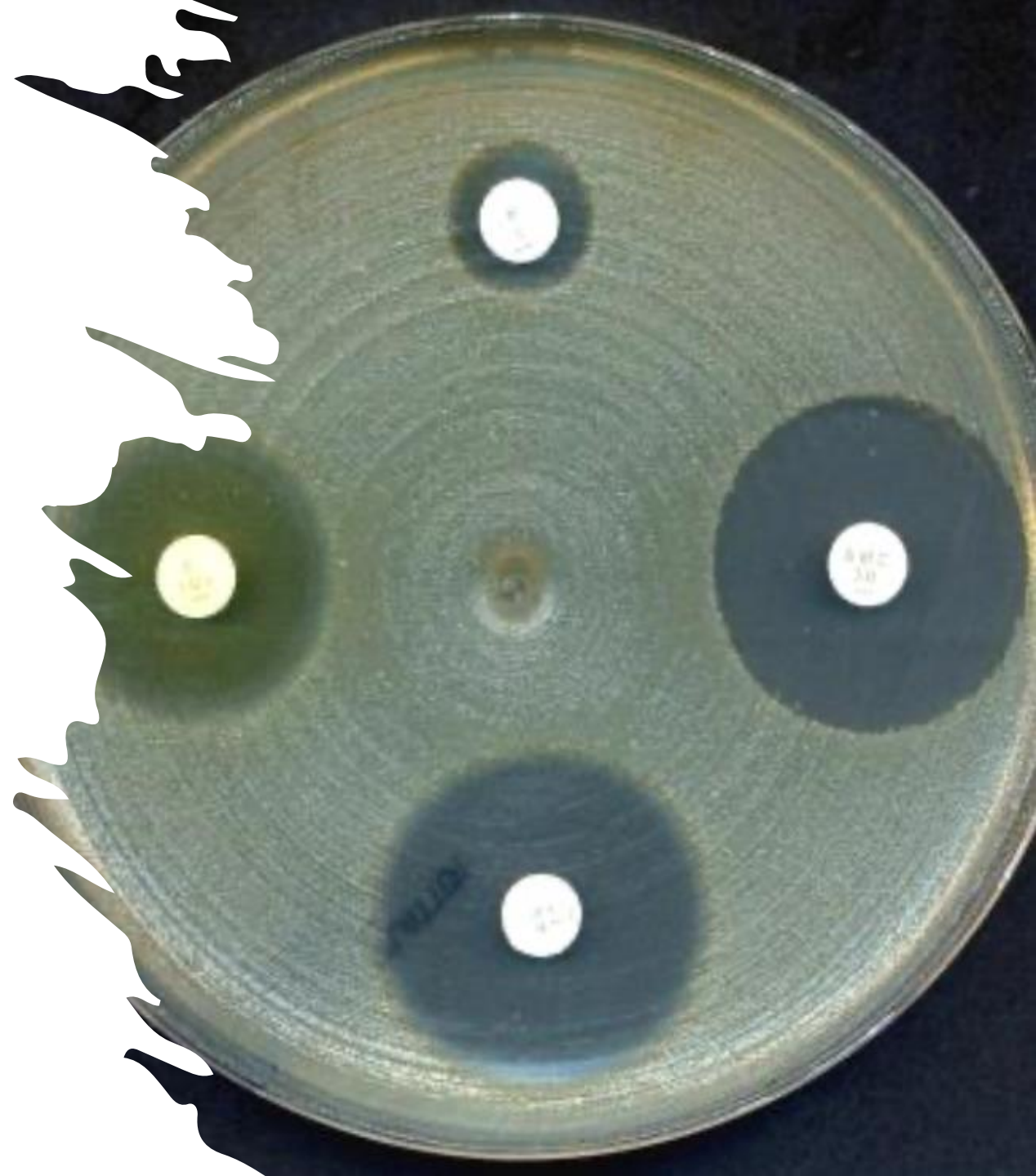
Enterobacterales –PİP-TAZ
S. aureus, *S. pneumoniae* - DA



Gram negatif HADT*

- PİP-TAZ 30/6 $\mu\text{g/ml}$
- CEFOTAKSİM 5 $\mu\text{g/ml}$
- CEFTAZİDİM 10 $\mu\text{g/ml}$
- MEROPENEM 10 $\mu\text{g/ml}$
- CİPROFLOKSASİN 5 $\mu\text{g/ml}$
- CZA10/4 $\mu\text{g/ml}$

YBÜ, Dahiliye, Onkoloji servisleri, iletişim





H-ID ve HADT sonuçları ile hastalara yapılan öneriler ve değerlendirme yapılsaydı aynı gün yapılabilecek potansiyel tanı ve tedavi önerileri

	YAPILAN ÖNERİLER	DEĞERLENDİRME YAPILSAYDI POTANSİYEL ÖNERİLER
Antibiyotik tedavisi etki kapsamının genişletilmesi (eskalasyon)	19	3
Antibiyotik tedavisi etki kapsamının daraltılması (de-eskalasyon)	2	4
Santral venöz kateter çekilmesi	6	1
Cilt flora kontaminasyonu	17	23
Ekokardiyografi	1	-

SONUÇ

- Rutin olarak kullanılan yöntemler hala kültüre bağlıdır = hızlı sonuçlar elde etmede darboğaz
- EUCAST HADT, hem hız hem de doğruluk açısından yeni ADT yöntemlerinin rekabet etmesini zorlaştıran yeni bir standart belirlemiştir-MONOMİKROBİYAL-TÜR
- Hedefe yönelik genotipik yöntemler belirli durumlarda (örn. yerel epidemiyoloji, hastane salgınları, yüksek riskli hastalar) gerekebilir.
- Hızlı fenotipik yöntemler – farklı tespit yaklaşımları, bazı yöntemler umut verici, ancak mevcut tüm ilaç/bakteri kombinasyonları için geçerliliğini doğrulamak çok zor.

Thank You!

